

## **Supplementary data**

**Journal of Bacteriology JB00370-19**

### **Methylation warfare: interaction of pneumococcal bacteriophages with their host**

Leonardo Furi<sup>1</sup>, Liam A. Crawford<sup>1</sup>, Guillermo Rangel-Pineros<sup>1</sup>, Ana Sousa Manso<sup>1</sup>, Megan De Ste Croix<sup>1</sup>, Richard D. Haigh<sup>1</sup>, Min J Kwun<sup>2</sup>, Kristine Engelsen Fjelland<sup>3&</sup>, Gregor Duncan Gilfillan<sup>3</sup>, Stephen D. Bentley<sup>4</sup>, Nicholas J. Croucher<sup>2</sup>, Martha R. Clokie<sup>1</sup>, Marco R. Oggioni<sup>1,\*</sup>

<sup>1</sup> Department of Genetics and Genome Biology, University of Leicester, Leicester, UK

<sup>2</sup> MRC Centre for Global Infectious Disease Analysis, Department of Infectious Disease Epidemiology, Imperial College London, London, United Kingdom.

<sup>3</sup> Department of Medical Genetics, Oslo University Hospital and University of Oslo, Oslo, Norway

<sup>4</sup> Parasites and Microbes, Wellcome Trust Sanger Institute, Hinxton, United Kingdom.

\*Correspondence to: Marco R Oggioni. Department of Genetics and Genome Biology, University of Leicester, University Road, Leicester, LE1 7RH, UK. Phone. +44 (0)116 252 2261, Email: [mro5@leicester.ac.uk](mailto:mro5@leicester.ac.uk)

**Table S1.** Bacterial strains and primers

Name	Background	Knock out	Relevant properties <sup>a</sup>	Reference
D39	-	-	Type 2 Avery's strain	(3, 27)
DP1004	Rx1	-	D39 unencapsulated derivative; Sm <sup>r</sup>	(27, 59)
FP486	D39	SPD_0449-455	<i>spnD39III</i> deleted mutant; Sp <sup>r</sup>	This work
FP470	DP1004	SPD_0449-455	<i>spnDP1004III</i> deleted mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
FP422	DP1004	SPD_0450-452	<i>spnDP1004IIIB</i> locked mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
FP423	DP1004	SPD_0450-452	<i>spnDP1004IIIC</i> locked mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
FP424	DP1004	SPD_0450-452	<i>spnDP1004IIIA</i> locked mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
FP425	DP1004	SPD_0450-452	<i>spnDP1004IIID</i> locked mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
MRO557	DP1004	SPD_0450-452	<i>spnDP1004IIIE</i> locked mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
MRO558	DP1004	SPD_0450-452	<i>spnDP1004IIIF</i> locked mutant; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
MRO587	FP470	-	SpSL1 prophage carrying strain	This work
MRO575	FP424	SPD_0450-452; SPD_455	<i>spnDP1004IIIA</i> locked::hsdR frameshifted; Sp <sup>r</sup> ; Sm <sup>r</sup>	This work
MRO594	FP424	SPD_0450-452; SPD_0930-1	<i>spnDP1004IIIA</i> locked::pezAT deleted mutant; Sp <sup>r</sup> ; Km <sup>r</sup>	This work
MRO598	FP424	SPD_0450-452; SPD_1081-2	<i>spnDP1004IIIA</i> locked::relBE2 deleted mutant; Sp <sup>r</sup> ; Cm <sup>r</sup>	This work
MRO602	FP424	SPD_0450-452; SPD_1550-1	<i>spnDP1004IIIB</i> locked::yefM-yoeB deleted mutant; Sp <sup>r</sup> ; Km <sup>r</sup>	This work
MRO589	FP422	SPD_0450-452; SPD_1108-9	<i>spnDP1004IIIB</i> locked::spnDP1004mcrBC deleted mutant; Sp <sup>r</sup> ; Sm <sup>s</sup> ; Km <sup>r</sup>	This work
MRO590	FP424	SPD_0450-452; SPD_1108-9	<i>spnDP1004IIIA</i> locked::spnDP1004mcrBC deleted mutant; Sp <sup>r</sup> ; Sm <sup>s</sup> ; Km <sup>r</sup>	This work
MRO628	FP424	SPD_0450-452; SPD_1737	<i>spnDP1004IIIA</i> locked::lytA deleted mutant; Sp <sup>r</sup> ; Sm <sup>s</sup> ; Km <sup>r</sup>	This work
MRO629	FP470	SPD_0449-445; SPD1259-60	<i>spnDP1004III</i> deleted mutant::spnDP1004I deleted mutant; Sp <sup>r</sup> ; Sm <sup>s</sup> ; Km <sup>r</sup>	This work
R6x <i>Δivr</i> <i>tvr</i> ::Janus	R6x	-	R6x <i>Δivr</i> <i>tvr</i> ::Janus; Km <sup>r</sup>	(38)
R6x <i>Δivr</i> <i>hsds</i> :: <i>tvr<sub>R</sub></i> <i>MV<sub>5</sub>ΔtvrR</i>	R6x	-	R6x <i>Δivr</i> <i>hsdS</i> :: <i>tvr<sub>R-MV5</sub>ΔtvrR</i>	(38)
Target	Template GenBank ID		Sequence (5' - 3')	
IF100	<i>Sp cassette</i>	KJ955483	GCTCTAGAACTAGTGGATC	(46)
IF101	<i>Sp cassette</i>	KJ955483	TTCCCTTCAAGAGCGATAC	(46)
LG70	<i>spnD39III</i>	NC_008533	GCAGTCTAACGCCATCAAATAC	(46)
LG71	<i>spnD39III</i>	NC_008533	GATCCACTAGTTCTAGAGCTTCGCTGTATTGTT CATC	(46)
ASM	<i>spnD39III</i>	NC_008533	GTATCGCTCTGAAGGGAGGCCATCGTTGGCTACT AAGATGT	This work
ASM	<i>spnD39III</i>	NC_008533	AGCATATCGTTACGAAGAACTTT	This work
LF83	SpSL1	KM882824	GTCACAAACACAAAAACTTCCAGAGT	This work
LF84	SpSL1	KM882824	ACCATATCGAGCCAACCTCTCCT	This work
LF88	SpSL1	KM882824	ACGGGCTATACTGCATCTTAGAGA	This work
LF89	SpSL1	KM882824	CTACTTGACCAGTCAGGCAGAAAT	This work
LF90	<i>csRNA3</i>	NC_008533	AGTAGGCTCTGGTCGTGAACAA	This work
LF91	<i>csRNA5</i>	NC_008533	AGGAACCTGCACCTGTTATGAAGGT	This work
LF92	<i>csRNA2</i>	NC_008533	GTATAGGGATGACTGGATGAATCGT	This work
LF93	<i>csRNA4</i>	NC_008533	CAGTTGGACTGCGCTTGAATGA	This work
LF94	<i>csRNA3</i>	NC_008533	GAGCAATCTCAGCCTCTCTCAA	This work
LF94	<i>csRNA5</i>	NC_008533	GCGCGACCAATCGTAAAC	This work
LF96	<i>csRNA2</i>	NC_008533	GATTAAGAGAACGTTAGGGATTCTATCA	This work
LF97	<i>csRNA4</i>	NC_008533	CTCTACTTCCACAGCCTTATCTTC	This work
LF106	<i>spnMcrBC</i>	NC_008533	CTTGTATCCGCTCTGCACG	This work

LF107	<i>spnMcrBC</i>	NC_008533	ATCAAACGGATCCCCAGCTTGTGAGATAGGAAACCTA GCAGGT	This work
LF108	<i>spnMcrBC</i>	NC_008533	CGCCTACGAGGAATTGTATCCCCAGTGTAGGAGCT TTAATATC	This work
LF109	<i>spnMcrBC</i>	NC_008533	ATCCTTCCTTGAAACCAGAAG	This work
LF149	<i>lytA</i>	NC_008533	CTTAATCCATTCTGTGATATTCC	This work
LF150	<i>lytA</i>	NC_008533	ATATCACATTATCCATTAAAATCAAACCTTCCTTCA ATGTTGCTACTATATT	This work
LF151	<i>lytA</i>	NC_008533	GCATAAGGAAAGGGCCATCCAGAACAGAAACTCCT	This work
LF152	<i>lytA</i>	NC_008533	TCTTATCTGGGTTGCCATGA	This work
hsdRU2	<i>hsdR</i>	NC_008533	CAAAATTAGAAAACATGCCTAGCG	This work
janus-hsdRR	<i>hsdR</i>	NC_008533	ATCAAACGGTCATATTGAACCCCTCCGCATTC	This work
hsdRd	<i>hsdR</i>	NC_008533	CAAACCAGCTTTCGAAGGCG	This work
janus-hsdRF	<i>hsdR</i>	NC_008533	AGGGATCCCGAGAAAGAAATGAGTGAGGAGG	This work
hsdRframef	<i>hsdR</i>	NC_008533	TTGAGTTATAAGTCTAAGAAAGAAATGAGTG	This work
hsdRframeler	<i>hsdR</i>	NC_008533	CTTTCTTAGACTTAAACTCAACCATTGAACC	This work
hsdR-janusF	<i>kan-rpsL</i>	(71)	GTTCAATATGACCGTTGATTTTAATGGATAATGTG	This work
hsdR-janusR	<i>kan-rpsL</i>	(71)	ATTCCTTCTCGGGATCCCTTCCTTATGC	This work

<sup>a</sup> Sm, streptomycin; Sp, spectinomycin; Km, kanamycin, Cm, chloramphenicol.

**Table S2.** Phage and host proteins identified with LC-MS/MS

Phage CDS	Function	GenBank ID	Molecular weight (kDa)	Total unique peptide count*
Cds32	portal protein	723007184	44	14
Cds33	prohead maturation protease	723007185	20	3
Cds34	capsid protein	723007186	47	17
Cds35	hypothetical protein	723007187	13	2
Cds36	head-tail adapter protein	723007188	12	3
Cds37	hypothetical protein	723007189	14	4
Cds39	major tail protein	723007191	24	20
Cds42	tail length tape-measure protein	723007194	98	16
Cds43	tail protein	723007231	27	10
Cds44	tail fiber PblB-like protein	723007195	179	43
Cds47	holin	723007198	11	3
Cds49	N-acetylmuramoyl-L-alanine amidase	723007200	36	3
Cds21	hypothetical protein	723007222	28	12
Cds24	hypothetical protein	723007225	10	2

\* Total number of different amino acid sequences that are associated only with this protein. Scaffold's calculated probability of correct protein identification threshold was set to >95%. With these settings the probability that a protein is correctly identified with a single peptide is of ~50%, while it increases with more uniquely associated peptides (e.g. to ~99.75% with two unique peptides).

**Table S3.** Time course of host differentially regulated genes during SpSL1 lytic infection

Gene D39 <sup>a</sup>	Name	Fold change <sup>b</sup>			Function	Lowes q-value
		10	50	90		
SPD_0001	<i>dnaA</i>	1.33	2.28	2.72	chromosomal replication initiator protein DnaA	2.20E-22
SPD_0039		0.66	0.90	0.48	conserved hypothetical protein	4.86E-27
SPD_0063	<i>strH</i>	1.15	0.13	0.06	beta-N-acetylhexosaminidase	0.00E+00
SPD_0066		1.46	0.51	0.46	PTS system, IIB component	1.03E-43
SPD_0068		1.86	0.52	0.47	PTS system, IID component	1.37E-83
SPD_0069		1.95	0.41	0.60	PTS system, IIA component	6.02E-140
SPD_0070	<i>agaS</i>	2.09	0.55	0.52	sugar isomerase domain protein AgaS	6.00E-107
SPD_0071	<i>galM</i>	2.48	0.52	0.57	aldose 1-epimerase	3.22E-124
SPD_0081		0.70	0.58	0.47	DNA-binding response regulator	4.01E-22
SPD_0088		1.84	0.34	0.22	ABC transporter, permease protein	0.00E+00
SPD_0088		1.84	0.34	0.22	ABC transporter, permease protein	0.00E+00
SPD_0089		2.32	0.34	0.19	ABC transporter, permease protein	0.00E+00
SPD_0089		2.32	0.34	0.19	ABC transporter, permease protein	0.00E+00
SPD_0090		1.90	0.24	0.12	ABC transporter, substrate-binding protein	0.00E+00
SPD_0092		1.38	0.45	0.33	conserved hypothetical protein	0.00E+00
SPD_0092		1.38	0.45	0.33	conserved hypothetical protein	0.00E+00
SPD_0104		0.73	0.43	0.31	LysM domain protein	6.83E-76
SPD_0118		1.82	2.00	1.00	conserved hypothetical protein	5.50E-06
SPD_0138		1.49	0.20	0.15	glycosyl transferase, group 1 family protein	0.00E+00
SPD_0139		1.60	0.16	0.14	glycosyl transferase, group 2 family protein	0.00E+00
SPD_0140		2.29	0.19	0.25	ABC transporter, ATP-binding protein	0.00E+00
SPD_0140		2.29	0.19	0.25	ABC transporter, ATP-binding protein	0.00E+00
SPD_0187	<i>nrdD</i>	1.77	18.23	42.50	anaerobic ribonucleoside-triphosphate reductase	0.00E+00
SPD_0188		1.81	20.28	44.72	conserved hypothetical protein	0.00E+00
SPD_0189		1.77	17.15	35.92	acetyltransferase, GNAT family protein	0.00E+00
					anaerobic ribonucleoside-triphosphate reductase activating	
SPD_0190	<i>nrdG</i>	1.92	18.31	47.31	protein	0.00E+00
SPD_0191		1.71	16.10	43.05	conserved hypothetical protein	0.00E+00
SPD_0247		1.27	0.25	0.17	glycosyl hydrolase, family protein 1	0.00E+00
SPD_0249		1.00	0.61	0.48	conserved hypothetical protein	3.32E-10
SPD_0261	<i>pepC</i>	0.62	0.69	0.49	aminopeptidase C	6.13E-33
					S-ribosylhomocysteine lyase (autoinducer-2 production	
SPD_0309	<i>luxS</i>	0.44	0.60	0.42	protein luxS) (AI-2 synthesis protein	7.14E-29
SPD_0310		0.66	0.63	0.46	conserved hypothetical protein	1.92E-51
SPD_0375	<i>serS</i>	0.74	0.54	0.48	seryl-tRNA synthetase	4.35E-18
SPD_0381	<i>acpP</i>	0.58	0.46	0.33	acyl carrier protein	2.94E-69
SPD_0386	<i>accB</i>	1.15	1.36	0.95	acetyl-CoA carboxylase, biotin carboxyl carrier protein	1.63E-04
SPD_0388	<i>accC</i>	1.20	1.36	1.03	acetyl-CoA carboxylase, biotin carboxylase	3.09E-04
SPD_0389	<i>accD</i>	1.12	1.37	0.82	acetyl-CoA carboxylase, carboxyl transferase, beta subunit	1.18E-14
SPD_0390	<i>accA</i>	1.12	1.37	1.23	acetyl-CoA carboxylase, carboxyl transferase, alpha subunit	1.05E-11
SPD_0391		1.02	1.34	1.53	conserved hypothetical protein	4.18E-17
SPD_0442	<i>pyrG</i>	0.61	0.32	0.29	CTP synthase	2.33E-106
SPD_0444		1.77	0.17	0.07	endo-beta-N-acetylglucosaminidase, putative	0.00E+00
SPD_0473	<i>blpY</i>	2.83	1.22	1.33	immunity protein BlpY	1.27E-21
SPD_0474		2.53	1.13	1.19	conserved hypothetical protein	7.85E-19
SPD_0546	<i>brnQ</i>	1.08	0.45	0.29	branched-chain amino acid transport system II carrier protein	3.51E-72
SPD_0575		0.68	0.63	0.39	sensor histidine kinase, putative	7.03E-27
SPD_0594		1.02	0.66	0.49	conserved hypothetical protein	8.22E-14
SPD_0608	<i>pyrF</i>	0.16	0.62	0.38	orotidine 5'-phosphate decarboxylase	9.31E-100
SPD_0609	<i>pyrE</i>	0.17	0.62	0.42	orotate phosphoribosyltransferase	2.48E-91
SPD_0612		2.50	3.36	2.71	lipoprotein, putative	1.32E-39
SPD_0614		2.07	2.43	2.36	ABC transporter, ATP-binding protein	4.12E-17
SPD_0632	<i>thiD</i>	0.71	0.55	0.49	phosphomethylpyrimidine kinase	8.96E-16
SPD_0636	<i>spxB</i>	0.70	0.44	0.23	pyruvate oxidase	2.57E-94
SPD_0637		0.55	0.43	0.26	glyoxalase family protein	3.02E-97
SPD_0641	<i>manA</i>	1.08	0.37	0.36	mannose-6-phosphate isomerase, class I	7.19E-222
SPD_0667	<i>sodA</i>	0.52	0.48	0.31	superoxide dismutase, manganese-dependent	1.24E-78
SPD_0731		0.19	0.56	0.49	DNA topology modulation protein FlaR, putative	1.10E-267

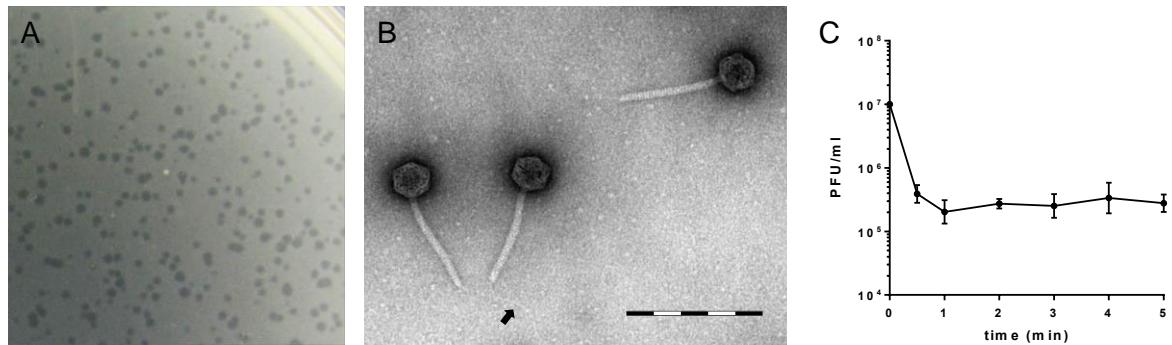
SPD_0732	<i>rpsT</i>	0.37	0.73	0.47	ribosomal protein S20	1.65E-42
SPD_0739		0.49	0.47	0.49	probable membrane lipoprotein TmpC precursor	1.86E-12
SPD_0762	<i>sufC</i>	0.72	0.52	0.45	FeS assembly ATPase SufC	5.73E-33
SPD_0763	<i>sufD</i>	0.79	0.56	0.46	FeS assembly protein SufD	7.07E-32
SPD_0764	<i>sufS</i>	0.91	0.56	0.43	cysteine desulfurases, SufS subfamily protein	1.47E-37
SPD_0765		0.97	0.49	0.41	SUF system FeS assembly protein, NifU family protein	4.48E-30
SPD_0799		0.63	0.67	0.43	conserved hypothetical protein	5.41E-10
SPD_0851	<i>pyrK</i>	0.20	0.60	0.30	dihydroorotate dehydrogenase electron transfer subunit	2.31E-97
SPD_0852	<i>pyrDb</i>	0.20	0.56	0.32	dihydroorotate dehydrogenase, catalytic subunit	1.01E-75
SPD_0853	<i>lytB</i>	0.48	0.68	0.49	endo-beta-N-acetylglucosaminidase precursor, putative	1.17E-10
SPD_0895	<i>hemH</i>	2.33	4.11	4.44	ferrochelatase	9.02E-86
SPD_0896	<i>mscL</i>	0.38	0.38	0.40	large conductance mechanosensitive channel protein MscL	4.15E-55
SPD_0898		0.63	0.35	0.32	membrane protein, putative	2.94E-44
SPD_0902	<i>trmE</i>	0.54	0.52	0.48	tRNA modification GTPase TrmE	2.09E-23
SPD_0903	<i>xylH</i>	0.45	1.13	0.56	4-oxaloacetate tautomerase	3.59E-12
SPD_0913		0.52	0.35	0.34	conserved hypothetical protein	2.47E-66
SPD_0987		0.66	0.64	0.48	conserved hypothetical protein TIGR00103	8.84E-14
SPD_1027		0.54	0.28	0.31	acetoin dehydrogenase, E1 component, beta subunit, putative	1.31E-60
SPD_1028	<i>acoA</i>	0.71	0.48	0.41	TPP-dependent acetoin dehydrogenase alpha-subunit	2.09E-23
SPD_1041	<i>nrdH</i>	0.89	4.17	6.35	glutaredoxin-like protein NrdH	7.95E-297
SPD_1042	<i>nrdE</i>	1.10	3.96	6.41	ribonucleoside-diphosphate reductase, alpha subunit	5.98E-159
SPD_1043	<i>nrdF</i>	1.10	3.56	6.91	ribonucleoside-diphosphate reductase, beta subunit	8.98E-252
SPD_1051	<i>lacC</i>	0.87	0.73	0.50	tagatose-6-phosphate kinase	1.19E-17
SPD_1052	<i>lacB</i>	0.79	0.63	0.38	galactose-6-phosphate isomerase, LacB subunit	6.28E-47
SPD_1053	<i>lacA</i>	0.79	0.61	0.48	galactose-6-phosphate isomerase, LacA subunit	1.83E-20
SPD_1075		2.26	2.04	2.61	transporter, FNT family protein, putative	5.08E-23
SPD_1078	<i>ldh</i>	0.53	0.50	0.26	L-lactate dehydrogenase	1.98E-135
SPD_1100	<i>zwf</i>	0.74	0.76	0.50	glucose-6-phosphate 1-dehydrogenase	7.32E-26
SPD_1107	<i>guoC</i>	0.45	0.60	0.86	guanosine monophosphate reductase	5.85E-28
SPD_1131	<i>carB</i>	0.24	0.83	0.59	carbamoyl-phosphate synthase, large subunit	2.82E-26
SPD_1132	<i>carA</i>	0.25	0.76	0.54	carbamoyl-phosphate synthase, small subunit	1.66E-29
SPD_1133	<i>pyrB</i>	0.21	0.65	0.45	aspartate carbamoyltransferase	2.56E-37
					pyrimidine operon regulatory protein/uracil	
SPD_1134	<i>pyrR</i>	0.20	0.52	0.41	phosphoribosyltransferase	1.48E-68
SPD_1141	<i>uraA</i>	0.19	0.35	0.24	uracil-xanthine permease	3.86E-152
SPD_1158	<i>gdhA</i>	0.81	0.74	0.40	NADP-specific glutamate dehydrogenase	3.87E-35
SPD_1217		0.44	0.54	0.67	conserved hypothetical protein	7.15E-23
SPD_1242		0.83	0.78	0.50	conserved hypothetical protein	2.67E-20
SPD_1286		0.47	0.63	0.63	TrmH family RNA methyltransferase group 3	3.30E-25
SPD_1318	<i>tuf</i>	0.66	0.57	0.45	translation elongation factor Tu	1.19E-14
SPD_1327	<i>bta</i>	0.53	0.55	0.31	bacteriocin transport accessory protein	1.37E-52
SPD_1334	<i>atpC</i>	1.15	0.72	0.38	ATP synthase F1, epsilon subunit	8.93E-45
SPD_1337	<i>atpA</i>	1.01	0.66	0.48	ATP synthase F1, alpha subunit	4.88E-29
SPD_1338	<i>atpH</i>	0.84	0.51	0.42	ATP synthase F1, delta subunit	1.05E-23
SPD_1340	<i>atpB</i>	0.80	0.71	0.49	ATP synthase F0, A subunit	2.67E-26
SPD_1360		0.60	0.55	0.34	conserved hypothetical protein	7.91E-43
SPD_1378		0.50	0.71	0.65	conserved hypothetical protein	1.01E-13
SPD_1402		0.78	0.43	0.38	non-heme iron-containing ferritin	1.63E-53
SPD_1413		0.95	0.44	0.82	ATP-dependent RNA helicase, putative	1.33E-30
SPD_1415		0.46	0.39	0.17	oxidoreductase, pyridine nucleotide-disulfide, class I	2.06E-164
SPD_1418	<i>pepQ</i>	0.59	0.54	0.32	proline dipeptidase PepQ	7.81E-75
SPD_1419		0.44	0.43	0.32	conserved hypothetical protein	5.92E-19
SPD_1439	<i>rpsO</i>	0.63	0.65	0.49	ribosomal protein S15	3.03E-17
SPD_1461	<i>psaB</i>	0.51	0.40	0.24	manganese ABC transporter, ATP-binding protein	6.36E-87
SPD_1462	<i>psaC</i>	0.67	0.45	0.29	manganese ABC transporter, permease protein, putative	1.51E-53
SPD_1463		0.70	0.51	0.40	ABC transporter, substrate binding lipoprotein	1.10E-46
SPD_1464	<i>psaD</i>	0.60	0.31	0.18	thiol peroxidase	8.68E-200
SPD_1467		0.31	0.79	0.54	conserved hypothetical protein	1.59E-19
SPD_1524		0.88	0.36	0.29	transcriptional regulator, GntR family protein	1.23E-53
SPD_1525		0.91	0.32	0.31	ABC transporter, ATP-binding protein	1.87E-71
SPD_1526		1.03	0.32	0.29	membrane protein, putative	5.69E-74
SPD_1527		1.06	0.73	0.50	membrane protein, putative	1.53E-33
SPD_1531	<i>scrK</i>	1.18	0.27	0.11	fructokinase	0.00E+00

SPD_1588		1.02	0.46	0.31	conserved hypothetical protein	2.03E-53
SPD_1589		0.89	0.38	0.27	lipoprotein, putative	1.83E-53
SPD_1590		1.07	0.57	0.33	general stress protein 24, putative	1.26E-59
SPD_1591		1.02	0.73	0.37	conserved hypothetical protein	1.41E-40
SPD_1594		0.80	2.16	4.16	transcriptional regulator	0.00E+00
SPD_1595		0.66	2.64	4.11	conserved hypothetical protein choline transporter (glycine betaine transport system)	0.00E+00
SPD_1642	<i>proW</i>	1.10	0.68	0.43	permease protein)	2.06E-32
SPD_1643	<i>proV</i>	0.88	0.59	0.39	choline transporter	2.61E-32
SPD_1644		0.84	0.53	0.40	conserved hypothetical protein	1.52E-27
SPD_1645		0.78	0.54	0.41	transcriptional regulator, MarR family protein	4.75E-17
SPD_1789		0.27	0.84	0.82	cell wall surface anchor family protein	1.92E-83
SPD_1795		0.15	0.44	0.43	conserved hypothetical protein	3.82E-111
SPD_1799		1.58	1.83	2.50	sensor histidine kinase, putative	3.31E-18
SPD_1823	<i>gap</i>	0.79	0.63	0.46	glyceraldehyde-3-phosphate dehydrogenase, type I	6.08E-16
SPD_1830		2.81	0.48	0.30	glycosyl hydrolase, family protein 1	0.00E+00
SPD_1830		2.81	0.48	0.30	glycosyl hydrolase, family protein 1	0.00E+00
SPD_1831		4.13	0.58	0.29	PTS system, IIC component	0.00E+00
SPD_1831		4.13	0.58	0.29	PTS system, IIC component	0.00E+00
SPD_1832		4.93	0.77	0.63	PTS system, IIB component	6.30E-215
SPD_1833		4.08	0.92	0.56	PTS system, IIA component	2.52E-136
SPD_1838	<i>yajC2</i>	0.48	0.90	0.66	preprotein translocase, YajC subunit	2.67E-13
SPD_1839	<i>tkt</i>	0.61	0.64	0.47	transketolase	2.20E-28
SPD_1865		2.09	1.98	1.31	alcohol dehydrogenase, zinc-containing	5.67E-27
SPD_1868	<i>tgt</i>	0.61	0.39	0.29	queuine tRNA-ribosyltransferase	4.24E-71
SPD_1874		0.76	0.21	0.17	LysM domain protein	0.00E+00
SPD_1895		0.48	0.78	0.61	cytidine/deoxycytidylate deaminase family protein	1.49E-25
SPD_1896	<i>gltX</i>	0.48	0.67	0.54	glutamyl-tRNA synthetase	6.02E-20
SPD_1968		2.26	0.12	0.08	conserved hypothetical protein	0.00E+00
SPD_1968		2.26	0.12	0.08	conserved hypothetical protein	0.00E+00
SPD_1969		1.39	0.17	0.08	glycosyl hydrolase-related protein	0.00E+00
SPD_1970		1.48	0.17	0.08	ROK family protein	0.00E+00
SPD_1971		1.61	0.17	0.10	glycosyl hydrolase-related protein	0.00E+00
SPD_1972		1.70	0.21	0.11	conserved hypothetical protein	0.00E+00
SPD_1972		1.70	0.21	0.11	conserved hypothetical protein	0.00E+00
SPD_1973		1.07	0.17	0.14	alpha-1,2-mannosidase, putative	0.00E+00
SPD_1974		1.75	0.25	0.11	conserved hypothetical protein	0.00E+00
SPD_2009		1.66	2.84	1.72	conserved hypothetical protein	2.53E-27
SPD_2010		1.13	0.40	0.39	conserved hypothetical protein	5.77E-17
SPD_2011	<i>glpF</i>	1.07	0.29	0.32	glycerol uptake facilitator protein	5.63E-97
SPD_2012	<i>glpO</i>	0.77	0.22	0.25	alpha-glycerophosphate oxidase	5.97E-200
SPD_2013	<i>glpK</i>	0.59	0.33	0.31	glycerol kinase	4.86E-73
SPD_2014		0.87	0.54	0.42	conserved hypothetical protein	7.33E-36
SPD_2015	<i>hsfO</i>	0.62	0.50	0.46	chaperonin, 33 kDa	5.16E-30
SPD_2016		0.71	0.47	0.43	TIM-barrel protein, nifR3 family protein, putative	8.68E-31
SPD_2046		2.54	1.69	1.92	cobalt ABC transporter, permease protein	9.75E-19
SPD_2053		0.44	0.85	0.78	conserved hypothetical protein	4.36E-16

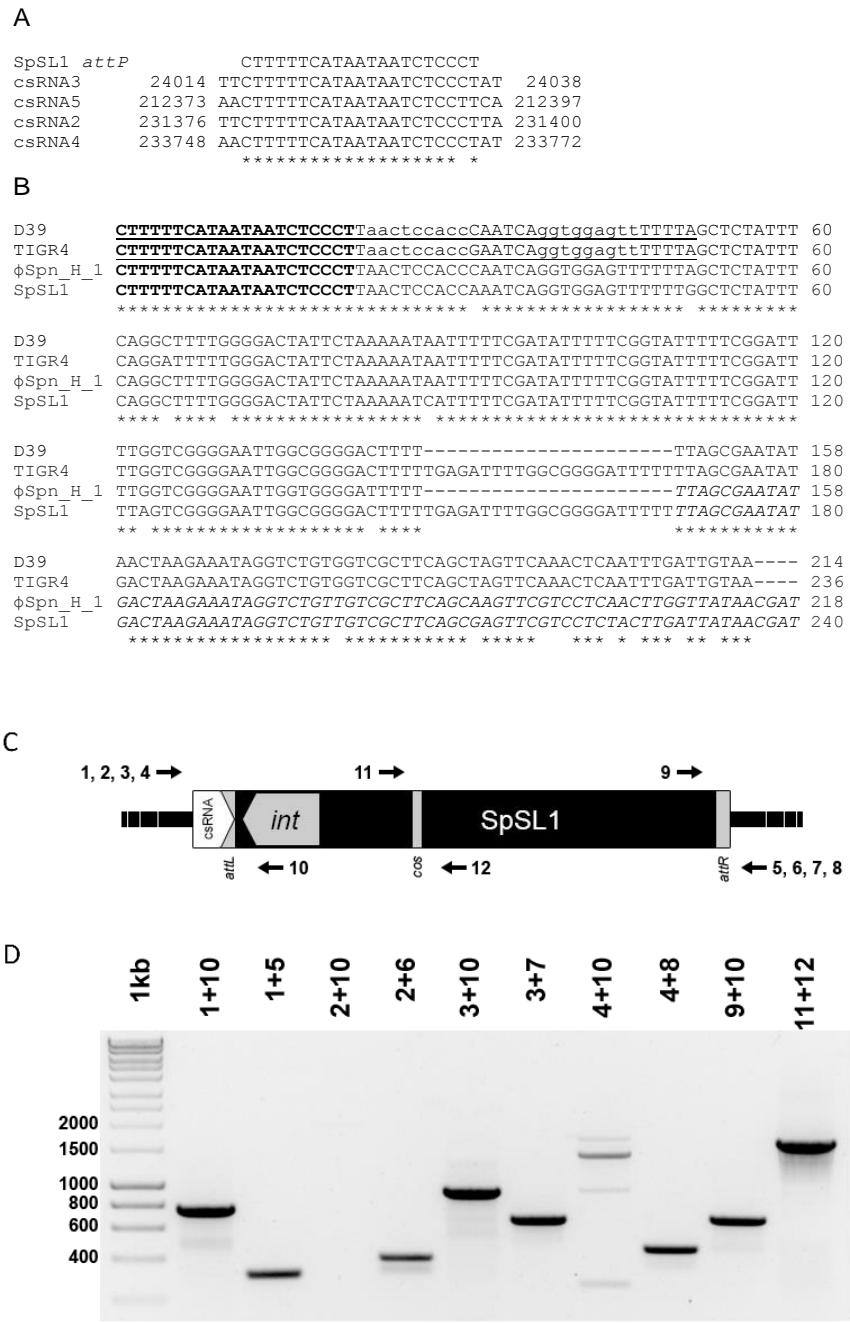
<sup>a</sup> Gene names refer to *S. pneumoniae* D39 genome (NC\_008533). <sup>b</sup> Fold change following gene expression variations after 10, 50, and 90 minutes post phage infection with respect to non-infected cells.

**Table S4.** Operon structure of the SpSL1 bacteriophage highlighting the CDSs and their predicted product expression time based upon RNA-Seq data.

CDS	Protein	Expression
<i>cds30</i>	hypothetical protein	Late
<i>cds31</i>	terminase, large subunit	Late
<i>cds32</i>	portal protein	Late
<i>cds33</i>	prohead maturation protease	Late
<i>cds34</i>	capsid protein	Late
<i>cds35</i>	hypothetical protein	Late
<i>cds36</i>	head-tail adapter protein	Late
<i>cds37</i>	hypothetical protein	Late
<i>cds38</i>	putative tail-component	Late
<i>cds39</i>	major tail protein	Late
<i>cds40</i>	hypothetical protein	Late
<i>cds41</i>	hypothetical protein	Late
<i>cds42</i>	tail length tape-measure protein	Late
<i>cds43</i>	tail protein	Late
<i>cds44</i>	tail fiber PblB-like protein	Late
<i>cds45</i>	hypothetical protein	Late
<i>cds46</i>	hypothetical protein	Late
<i>cds47</i>	holin	Late
<i>cds48</i>	holin 1	Late
<i>cds49</i>	N-acetylmuramoyl-L-alanine amidase	Late
<i>cds50</i>	hypothetical protein	hypothetical
<i>cds01</i>	integrase	Lysogeny
<i>cds02</i>	hypothetical protein	Lysogeny
<i>cds03</i>	hypothetical protein	Lysogeny
<i>cds04</i>	CI repressor	Lysogeny
<i>cds05</i>	hypothetical protein	Early
<i>cds06</i>	hypothetical protein	Early
<i>cds08</i>	prophage antirepressor protein	Early
<i>cds09</i>	hypothetical protein	Early
<i>cds10</i>	hypothetical protein	Early
<i>cds11</i>	DnaD	Early
<i>cds12</i>	hypothetical protein	Early
<i>cds13</i>	DnaC	Early
<i>cds14</i>	hypothetical protein	Early
<i>cds15</i>	C-5 cytosine-specific DNA methylase	Early
<i>cds16</i>	N-6 adenine-specific DNA methylase	Early
<i>cds17</i>	DNA-binding protein	Early
<i>cds18</i>	hypothetical protein	Early
<i>cds19</i>	hypothetical protein	Early
<i>cds20</i>	hypothetical protein	Early
<i>cds21</i>	hypothetical protein	Early
<i>cds22</i>	hypothetical protein	Early
<i>cds23</i>	hypothetical protein	Early
<i>cds24</i>	hypothetical protein	Early
<i>cds25</i>	Cro repressor	Early
<i>cds26</i>	hypothetical protein	Early
<i>cds27</i>	site-specific recombinase	Early
<i>cds28</i>	hypothetical protein	Very Late
<i>cds29</i>	endonuclease	Very Late

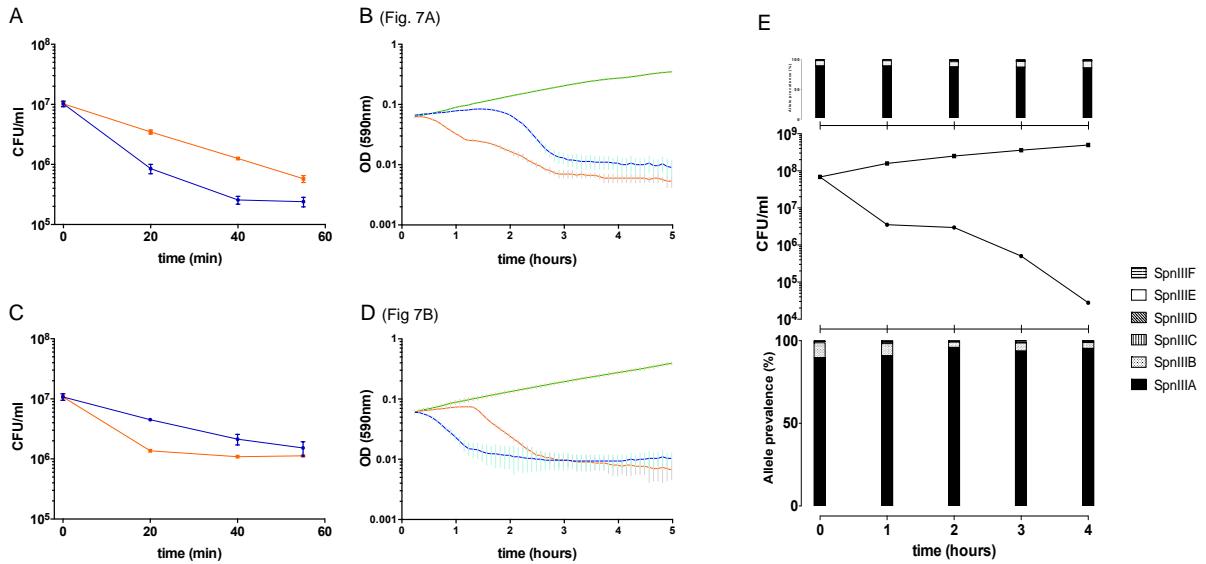


**Figure S1.** Plaques produced by SpSL1 on CAT-Galactose agar plates using a *spnDP1004/III* deletion mutant as bacterial cell lawn (panel A). Electron micrographs of purified SpSL1 showing *Siphoviridae* structure; arrow indicates the tail fiber. Bar 200 nm (panel B). Adsorption of SpSL1 to whole *spnDP1004/III* deleted mutant cells (panel C).

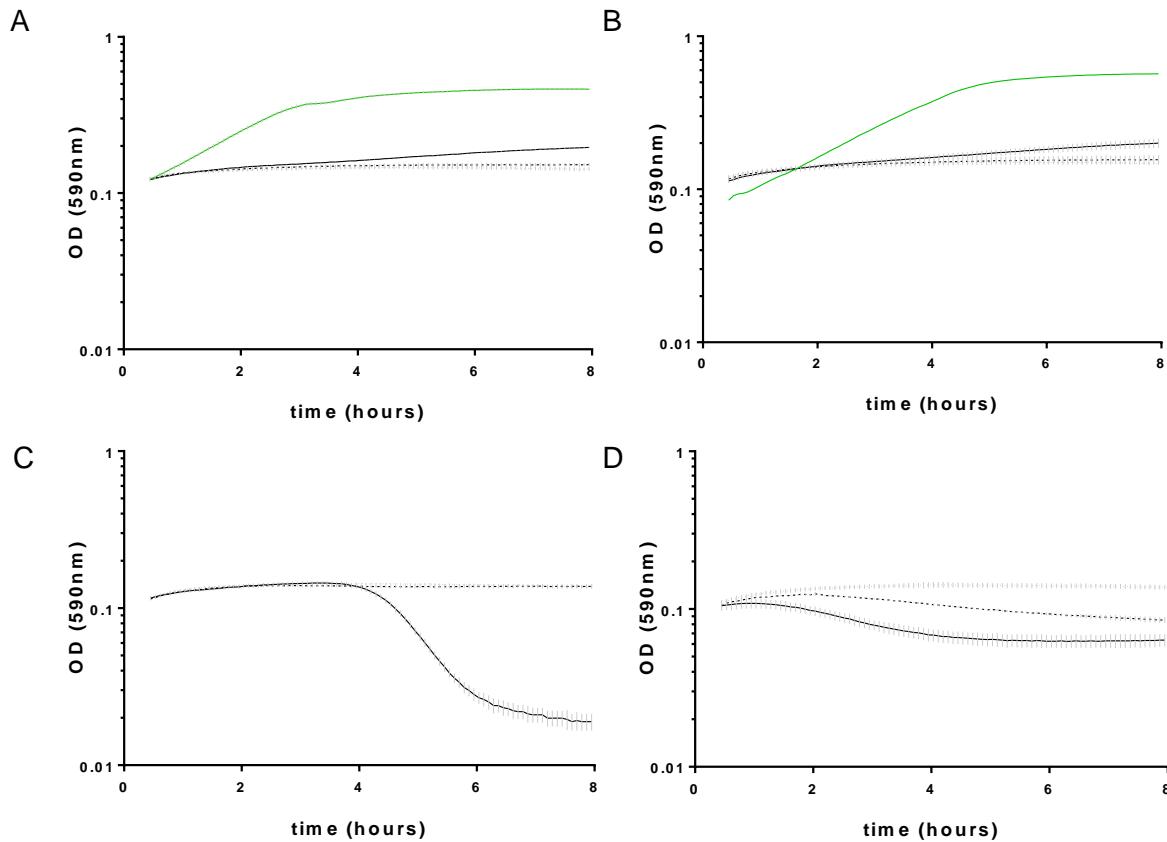


**Figure S2. Attachment site analysis.** Alignment of phage *att* sequence and complementary regions in the *S. pneumoniae* genome (panel A). The positions of the sequences belonging to the four csRNAs are reported based on D39 genome sequence (GenBank accession number NC\_008533). Comparison between the *S. pneumoniae* genome sequence following csRNA2 coding region and the sequence downstream of the *attP* in bacteriophages (panel B). The *attL* sequence is highlighted in bold. The final portion of the csRNA2 coding region is underlined and nucleotides forming the stem of the transcriptional terminator hairpin are shown in lowercase. The 5' phage integrase coding region is italicized. D39 and TIGR4 are representative of phage-free *S. pneumoniae* genome sequences, while φSpn\_H\_1 is one of the two prophages of Hungary19A-6 strain. Panel C shows the schematic map of lysogenic SpSL1. Internal cos site (red), integrase gene (grey), duplicated attachment sites (cyan) and a generic csRNA (orange) are showed. Primers marked 1-8 in the picture correspond to LF90-97 (Table S1), which are designed to specifically amplify the *S. pneumoniae* genome region around csRNA2, 3, 4, and 5. Primers labelled 9-12 coincide with LF89, 88, 83, and 84 (Table S1) respectively. Panel D shows PCR amplification of the four predicted attachment sites of a SpSL1 lysogenic (MRO587 mutant), with and without the integrated phage (well 1-8). Control PCRs for the identification of excised phage

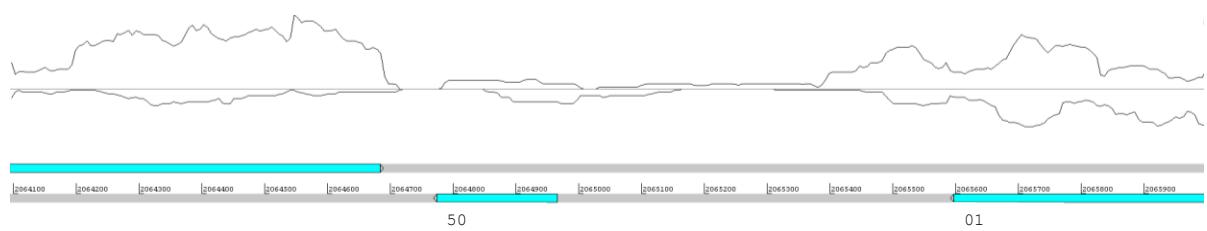
DNA and phage in the lysogenic form are in wells 9 and 10 respectively. Predicted band sizes from left to right are the following: 775, 319, 880, 408, 953, 692, 895, 469, 680, and 1638 base pairs. Note that the third (well 3) and fourth (well 7) samples are negative with the latter showing amplified fragments that are not of the expected size. DNA ladder sizes are reported in base pairs.



**Figure S3: Abortive infection associated phenotypes.** Data show the killing (left panels) in comparison to the culture turbidity (right panels; for comparative analysis panels of Fig.7 are reproduced here [Fig.S5B is Fig.7A and Fig.5D is Fig.7B]) of *spnDP1004IIIA* (panel A and B) and *spnDP1004IIB* (panel C and D) locked mutants following infection with SpnIIIA-methylated (blue) and SpnIIB-methylated (orange) SpSL1 phage (MOI=2). CFUs of survived bacteria were measured at 20, 40, and 55 minutes after the infection (panel A and C). OD<sub>590</sub> was read for 5 hours, with green line representing the non-infected controls (panel B and D). Panel E shows lack of any *spnIII* allele selection upon SpnIIIA-methylated phage infection (MOI=0.25). In the central panel CFU counts of the *wt* strain are shown (circles, phage infected strain; squares, negative control). Alleles in life bacteria were quantified for *s.spnDP1004III* variants after overnight incubation of an agar plate for each time point. SpnIII variants composition in the non-infected controls are shown above and of the infected cells below the growth curve. The patterns represent each allele conformation (*s.spnDP1004IIIA* black, *s.spnDP1004IIB* dots, *s.spnDP1004IIIC* vertical stripes, *s.spnDP1004IID* diagonal stripes, *s.spnDP1004IIIE* white, *s.spnDP1004IIIF* horizontal stripes).



**Figure S4:** The Abi mechanism requires the *de novo* synthesis of cellular constituents to produce cell death. Control experiments, using *spnDP1004IIIA* (panel A) and *spnDP1004IIB* (panel B) locked mutants, demonstrate the inhibition of cell replication in the presence of 4 mg/L (straight black line), 16 mg/L (dashed black line), and 64 mg/L (dotted black line) of chloramphenicol (Cm). The *spnDP1004IIIA* locked mutant was infected with SpnIIIA-methylated SpSL1 phage (MOI=2.5) in presence of similar Cm concentrations (panel C). Bacteriophage lytic cycle was delayed with 4 mg/L of Cm and was blocked at 16 mg/L (dashed line). When the *spnDP1004IIB* locked mutant was infected with SpnIIIA-methylated SpSL1 phage (MOI=2.5) the Abi phenotype was slower with 4 and 16 mg/L Cm and arrested in the presence of 64 mg/L Cm.



**Figure S5.** Zoomed in alignment of RNA-Seq of the lysogen at the hypothetical protein (*cds50*) in the centre, *cds49* (left) and *cds01* (right) highlighting the high degree of antisense RNA at *cds50* and throughout this region.